Response to Office Action dated August 22, 2006

Amendments to the Claims:

This listing of the claims will replace all prior versions, and listings, of claims in the application:

Listing of Claims:

- (original) An expression vector for producing IL-21 protein comprising the following operably linked elements:
 - (a) a prokaryotic origin of replication;
 - (b) a transcriptional initiation DNA element;
 - (c) a polynucleotide sequence as shown in SEQ ID NO:27; and
 - (d) a transcriptional terminator.
- $2. \qquad \hbox{(original) The expression vector of claim 1 which further comprises a selectable marker.}$
- (original) An expression vector comprising the pTAP337 vector, deposited with the American Type Culture Collection in Manassas, VA. under Patent Deposit Designation PTA-4853.
- (original) A prokaryotic host cell transformed with the expression vector according to claims 1, 2 or 3.
- 5. (original) The host cell of claim 4, wherein the host cell is $E.\ coli$ strain W3110.
- 6. (currently amended) A method for producing IL-21 proteins protein comprising:
- (a) culturing a host cell according to claim 5 in growth medium under conditions wherein IL-21 is expressed;
 - (b) recovering the host cells from the growth medium; and
 - (c) isolating the IL-21 protein from the host cells.
- 7. (with drawn) A method for producing ${\rm IL}\text{-}21$ proteins comprising:

Application Serial No.: 10/735,149

Amendment dated: December 20, 2006

Response to Office Action dated August 22, 2006

- (a) culturing a host cell according to claim 5 in growth medium by fed batch fermentation;
 - (b) recovering the host cells from the growth medium; and
 - (c) isolating the IL-21 protein from the host cells.
- 8. (withdrawn) A method for producing an IL-21 protein comprising:
- (a) culturing a host cell according to claim 4 or claim 5 in a shake flask to an OD600 of 5 to 20 in a growth medium;
- (b) inoculating a fermentation vessel with 1 to 12% v/v of shake flask medium containing host cells;
- (c) culturing the host cells in a growth medium at a pH of 6.2 to 7.2, wherein a feed solution is fed into the fermentation vessel before 15 hours elapsed fermentation time (EFT);
- (d) adding an inducing agent to the fermentation vessel at 20 to 30 hours EFT; and
 - (e) harvesting the host cells at 48 to 56 hours EFT.
- 9. (withdrawn) The method of claim 8, wherein the inducing agent is isopropyl thiogalactopyranoside (IPTG) at 0.5 to 2 mM.
- 10. (withdrawn) The method of claim 8, wherein the feed solution comprises a carbohydrate selected from the group consisting of glycerol and glucose at a concentration of growth medium, and a feed rate of 5-15 grams of carbohydrate per hour.
- 11. (withdrawn) The method of claim 10, wherein the glycerol is 40 to 70% v/y glycerol or the glucose is 40 to 70% w/v glucose.
- 12. (withdrawn) The method of claim 10, wherein the glycerol is about 70% v/v or the glucose is about 60% w/v.
- $\qquad \qquad 13. \qquad \text{(withdrawn)} \quad A \quad \text{method} \quad \text{of} \quad \text{producing} \quad \text{IL-}21 \quad \text{protein} \\ \text{comprising:} \quad$
- (a) seeding a flask with an inoculum comprising an E. coli W3110 host cell expressing an IL-21 polypeptide as shown in SEQ ID NO:28, or an E. coli W3110 host cell comprising pTAP337 vector wherein an IL-21

Application Serial No.: 10/735,149

and

Amendment dated: December 20, 2006

Response to Office Action dated August 22, 2006

polypeptide is expressed, and with growth medium comprising about 5 g/L glycerol;

- (b) culturing the inoculum in growth medium for 16-20 hours at about 30°C;
- (c) transferring the cultured inoculum in growth medium to a batch fermentor at a concentration of 0.5-5% v/v inoculum;
- (d) fermenting the batch fermentation at about 37° and about pH 6.8; with about 2% glycerol;
- (e) introducing a glucose feed at about 8 hours elapsed fermentation time (EFT) of about 9.5 g glucose/liter/hour and continuing until end of a fermentation run;
- (f) adding IPTG at about 24 hour EFT to final concentration of 0.5 to 2 mM;
 - (g) fermenting about 28 hours after addition of IPTG;
 - (h) harvesting fermentation broth from the fermenter;
 - adding an equal volume of water to the fermentation broth;
- (j) homogenizing and centrifuging the fermentation broth to collect a cell pellet or cell slurry comprising IL-21 protein material.
- 14. (withdrawn) A method for isolating insoluble IL-21 protein comprising a sequence of amino acid residues as shown in SEQ ID NO:28 comprising the steps of:
- (a) separating water insoluble IL-21 protein material from a cell pellet or cell slurry;
- (b) dissolving the insoluble IL-21 protein material in a chaotropic solvent;
- (c) diluting the chaotropic solvent and refolding the IL-21 protein; and
- (d) isolating the IL-21 protein, wherein the isolated IL-21 protein is capable of being biologically active.
- \$15.\$ (withdrawn) The method of claim 14 wherein the isolated IL-21 protein is at least 90% pure.

Response to Office Action dated August 22, 2006

16. (withdrawn) The method of claim 14 wherein the isolated IL-21 protein is at least 90% pure and has an endotoxin level of less than 10 endotoxin units per mg IL-21 protein.

- 17. (withdrawn) A method for isolating insoluble IL-21 protein comprising a sequence of amino acid residues as shown in SEQ ID NO:28 comprising the steps of:
- (a) separating from a fermentation broth a cell pellet or cell slurry comprising water insoluble IL-21 protein material;
- (b) homogenizing the cell pellet or cell slurry to collect inclusion bodies:
- (c) dissolving the insoluble IL-21 protein material in a chaotropic solvent comprising a guanidine salt;
- (d) diluting the chaotropic solvent by addition of a refolding buffer comprising arginine salts and a mixture of reducing and oxiding components;
- (e) isolating the IL-21 protein by removing unfolded and aggregated proteins by filtering; and
- (f) purifying the IL-21 refolded protein on a cation exchange column; wherein the isolated and purified IL-21 protein is capable of being biologically active.
- 18. (withdrawn) A method for isolating insoluble IL-21 protein comprising a sequence of amino acid residues as shown in SEQ ID NO:28 comprising the steps of:
- (a) separating from a fermentation broth a cell pellet or cell slurry comprising water insoluble IL-21 protein material;
- (b) homogenizing the cell pellet or cell slurry to collect inclusion bodies:
- (c) dissolving the insoluble IL-21 protein material in a chaotropic solvent comprising a guanidine salt; and
- (d) diluting the chaotropic solvent by addition of a refolding buffer comprising arginine salts and a mixture of reducing and oxidizing components;
- (e) isolating the IL-21 protein by removing unfolded and aggregated proteins by filtering;

Response to Office Action dated August 22, 2006

- (f) purifying the IL-21 refolded protein on a cation exchange column; and
- (g) purifying the IL-21 eluate from step (f) on a hydrophobic interaction column, wherein the isolated and purified IL-21 protein is capable of being biologically active.
- (withdrawn) A method for isolating insoluble IL-21 protein comprising a sequence of amino acid residues as shown in SEQ ID NO:28 comprising the steps of:
- (a) separating from a fermentation broth a cell pellet or cell slurry comprising water insoluble IL-21 protein material;
- (b) homogenizing the cell pellet or cell slurry to collect inclusion bodies;
- (c) dissolving the insoluble IL-21 protein in a chaotropic solvent comprising about 6M guanidine hydrochloride, 40 mM dithiothreitol (DTT) for about one hour at room temperature;
- (d) refolding the dissolved inclusion bodies in a solution by diluting into refolding buffer comprising about 0.75 M arginine, 2 mM DTT/4 mM cystine oxidation-reduction pair at least 20 times;
- (e) adjusting pH to about 5.5 with about 20% acetic and allowing the solution to react for at least five hours;
- $\mbox{(f)} \qquad \mbox{diluting the solution with about } 1 + 1.4 \mbox{ volumes } 25 \mbox{ mM} \\ \mbox{acetate, pH 5.5;} \\ \mbox{}$

(g) filtering the solution;

- (h) loading solution on resin column equilibrated to pH 5.5 using sodium acetate buffer:
- $\mbox{(i)} \qquad \mbox{washing the resin column with about } 0.4 \ \mbox{M} \ \mbox{sodium}$ chloride:
- (j) washing the resin column with about 0.75 M sodium chloride to elute bound IL-21 protein;
- (k) adding ammonium sulfate to a concentration of about 1.5
 M to eluate and filtering cluate solution;
- (1) loading eluate onto a Tosohaas butyl 650-M column equilibrated to 1.5 M ammonium sulfate, 0.05 M sodium chloride in sodium acetate buffer;
- (m) washing column with about 0.15 M ammonium sulfate, 0.05 sodium chloride in sodium acetate buffer;

Response to Office Action dated August 22, 2006

(n) diluting the eluate to a conductivity of about 30 mS/cm with water:

- (o) loading eluate onto a SP Sepharose HP column equilibrated with sodium acetate buffer:
- (p) washing column with 20-column volume linear gradient from 0.3 to 0.7 M sodium chloride;
 - (q) concentrating the IL-21 protein; and
- $\mbox{(r)} \qquad \mbox{exchanging buffer to formulation buffer using tangential} \label{eq:constraint} flow ultrafiltration.$
- (withdrawn) The method according to claims 13, 14, 15, or
 wherein biological activity is measured using a IL-21 receptor-binding cell assay.
- (withdrawn) A composition comprising an IL-21 protein comprising amino acids residues 1-163 as shown in SEQ ID NO:28 at a concentration of about 10 mg/ml IL-21 protein in about 10 mM histidine, 4.7% mannitol at pH 5.3
- 22. (currently amended) A host cell from a strain of zGOLD1, deposited with the American Type Culture Collection (ATCC) in Manassas, VA as <u>PTA-5698</u>, transformed with an expression vector comprising a pTAP337 vector, deposited with the ATCC under Patent Deposit Designation PTA-4853.